

PSY 511

Chemical communication

Rick Gilmore

2021-10-07 11:15:35

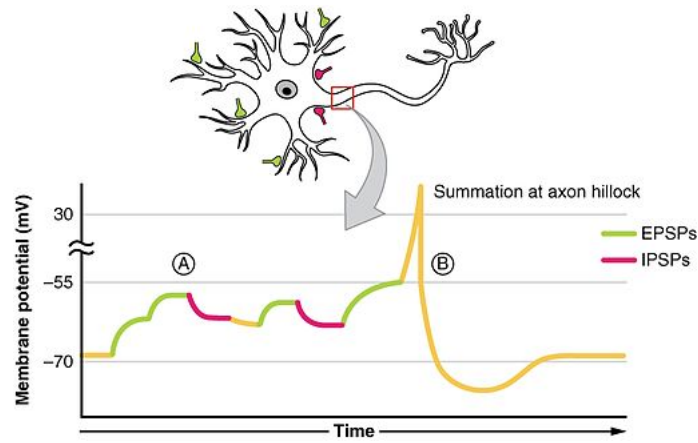
- Neural communication
 - What triggers the action potential?
 - Synaptic transmission
 - Synapse Types & Locations
 - Steps in chemical transmission
 - Receptor/channel types
 - Leak/passive
 - Transporters/exchangers
 - Ionotropic receptors (receptor + ion channel)
 - Metabotropic receptors (receptor only)
 - Receptors generate *postsynaptic potentials (PSPs)*
 - NTs inactivated
 - Questions to ponder
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 - What are they?
 - Things to know
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 - γ aminobutyric acid (GABA)
 - Other amino acid NTs
 - Acetylcholine (ACh)
 - Curare
 - Atropine
 - Monoamine NTs
 - Information processing
 - Dopamine
- {r, fig.cap="http://thebrain.mcgill.ca/flash/a/a_03/a_03_cl/a_03_cl_que/a_03_cl_que_01.html"}
 - Clinical relevance
 - Inactivated via
 - Norepinephrine
 - Clinical relevance
 - Inactivated by

- Adrenaline/Epinephrine
- Serotonin (5-hydroxytryptamine or 5-HT)
 - Clinical relevance
- Melatonin
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 - Thinking about neurochemical influences
- Enteric nervous system
 - Anatomy
 - Gut/brain connection
 - Physiology
- References

Neural communication

What triggers the action potential?

- Soma receives input from dendrites (and on soma directly)
- Axon hillock sums/integrates



Source:

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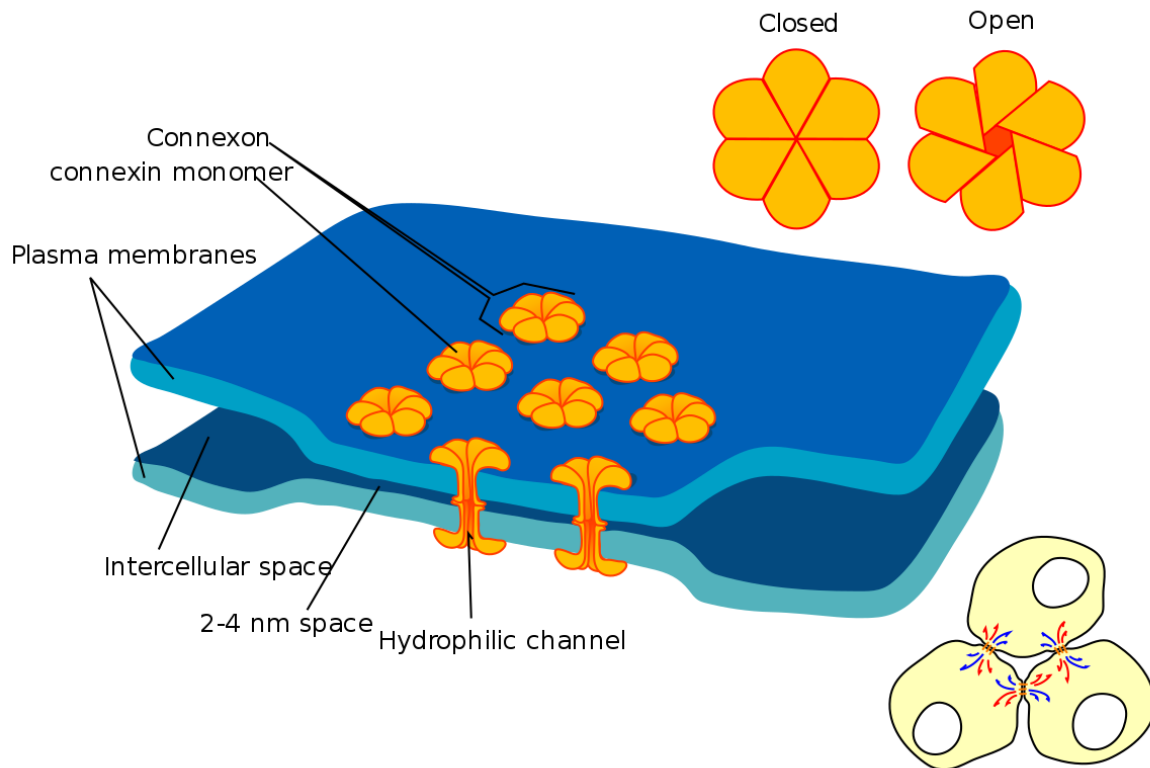
- If sum > threshold, action potential “fires”
- Action potential propagates along the axon
- Action potential’s rapid change in voltage triggers neurotransmitter (NT) release

Synaptic transmission

Synapse permits neuron to pass electrical or chemical messages to another neuron or target cell (muscle, gland, etc.)

Synapse Types & Locations

- Chemical
- Electrical
 - Gap junctions
 - *Cytosol* (and ionic current) flows through adjacent neurons

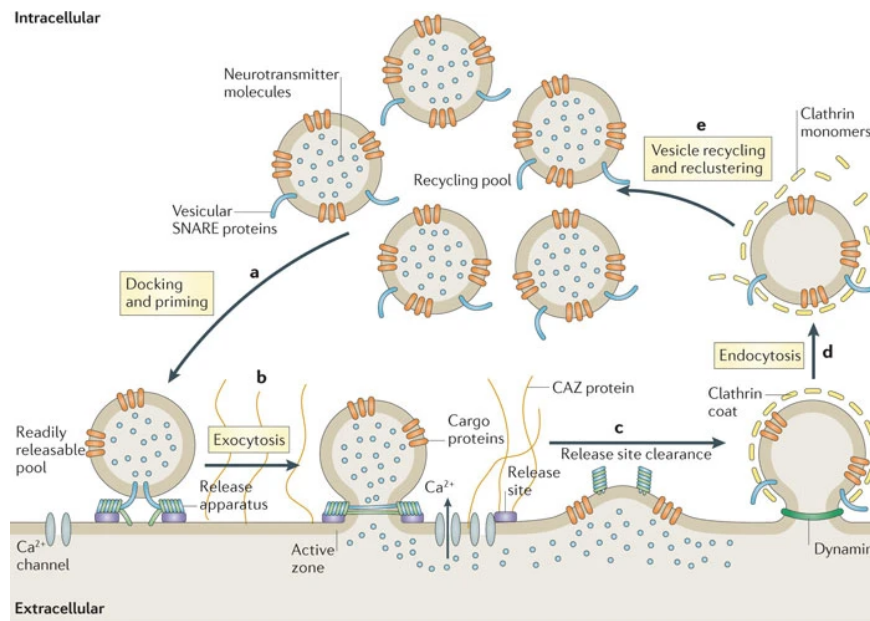


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By Mariana Ruiz [LadyofHats](//commons.wikimedia.org/wiki/User:LadyofHats "User:LadyofHats") - the diagram i made myself using the information on this websites as source: <http://academic.brooklyn.cuny.edu/biology/bio4fv/page/gap-junctions.html> (<http://academic.brooklyn.cuny.edu/biology/bio4fv/page/gap-junctions.html%22%3E>);[1] http://www-biology.ucsd.edu/classes/bipn140.FA05/10_2.jpg (http://www-biology.ucsd.edu/classes/bipn140.FA05/10_2.jpg%22%3E);[2] <http://www.colorado.edu/MCDB/MCDB1150/ohd/gapjunctionmodel.JPG> (<http://www.colorado.edu/MCDB/MCDB1150/ohd/gapjunctionmodel.JPG%22%3E>);[3] and <http://www.lrz-muenchen.de/~jmd/gap%20junction2.gif> (<http://www.lrz-muenchen.de/~jmd/gap%20junction2.gif%22%3E>);[4]. Made with Adobe Illustrator. Image renamed from [File:Gap_cell_junction.svg&action=edit&redlink=1](//commons.wikimedia.org/w/index.php?title=File:Gap_cell_junction.svg&action=edit&redlink=1) (File:Gap_cell_junction.svg&action=edit&redlink=1) class="new" title="File:Gap (File:Gap) cell junction.svg (page does not exist)">File:Gap (File:Gap) cell junction.svg, Public Domain, Link (<https://commons.wikimedia.org/w/index.php?curid=6027074>)

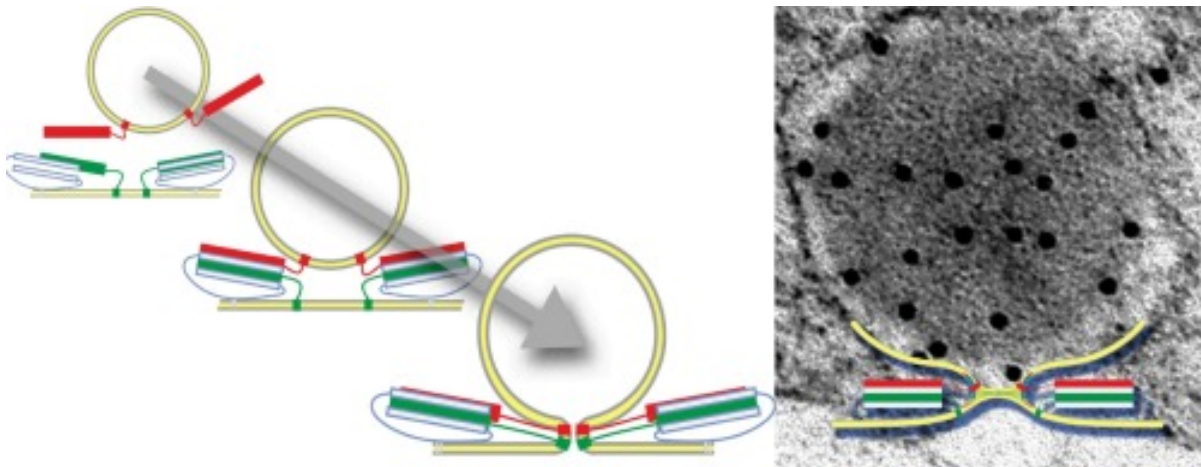
Steps in chemical transmission

- Voltage-gated calcium Ca^{++} channels open
- Ca^{++} influx causes synaptic vesicles to bind with presynaptic membrane, fuse with membrane, spill contents via exocytosis

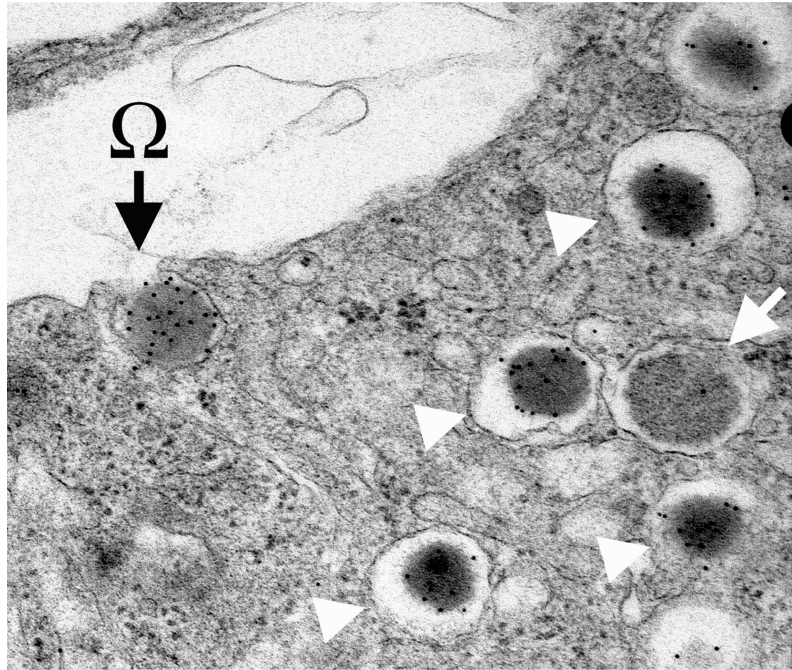


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(Hauke, Neher, & Sigrist, 2011) (<http://dx.doi.org/10.1038/nrn2948>)



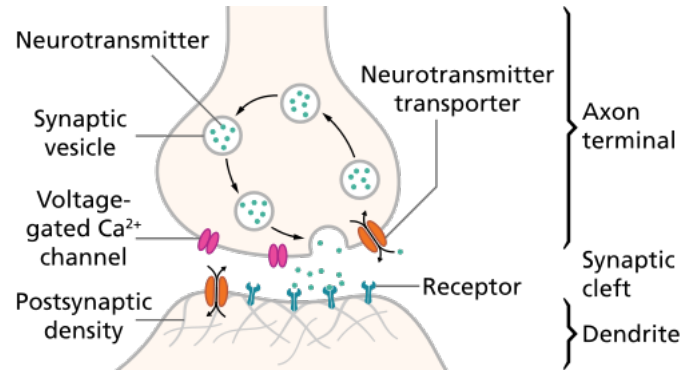
(Hastoy, Clark, Rorsman, & Lang, 2017) (<https://doi.org/10.1016/j.ceca.2017.10.005>)



(Hastoy, Clark, Rorsman, & Lang, 2017) (<https://doi.org/10.1016/j.ceca.2017.10.005>)



- NTs diffuse across *synaptic cleft*
- NTs bind with *receptors* on *postsynaptic membrane*
 - Cause some post-synaptic effect
- NTs unbind from receptor
- NTs inactivated
- NTs diffuse along concentration gradient



Source: https://commons.wikimedia.org/wiki/File%3ASynapseSchematic_en.svg
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Relative sizes

- Neural membrane ~8 nm
- Synaptic vesicles ~40-60 or ~90-120 nm
- Synaptic cleft ~20-50 nm
- Cleft small relative to vesicles

Receptor/channel types

Leak/passive

- Vary in selectivity, permeability

Transporters/exchangers

- Ionic
 - Na^+ / K^+ ATP-ase/pump
- Chemical
 - e.g., Dopamine transporter (DAT)

Ionotropic receptors (receptor + ion channel)

- Ligand-gated
- Open/close channel
- Ions flow in/out depending on membrane voltage and ion type
- Fast-responding (< 2 ms), but short-duration effects (< 100 ms)

Metabotropic receptors (receptor only)

- G-proteins ->
- Trigger 2nd messengers
- Open/close adjacent channels, change metabolism

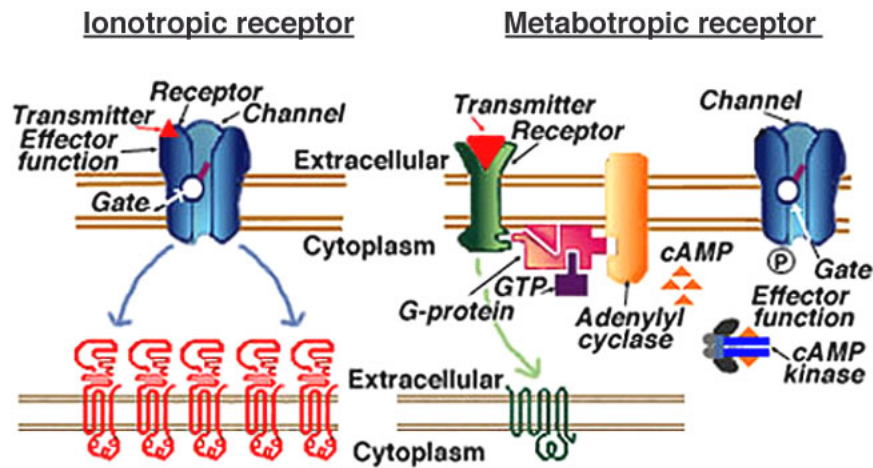


Fig. 5a. Ionotropic receptors and their associated ion channels form one complex (top). Each iGluR is formed from the co-assembly of multiple (4-5) subunits (From Kandel et al., 1991).

Fig. 5b. Metabotropic receptors are coupled to their associated ion channels by a second messenger cascade (top). Each mGluR is composed of one polypeptide, which is coupled to a G-protein (from Kandel et al., 1991).

Receptors generate postsynaptic potentials (PSPs)

- Small voltage changes
- Amplitude scales with # of receptors activated
- *Excitatory PSPs (EPSPs)*
 - Depolarize neuron (make more +)
- *Inhibitory (IPSPs)*
 - Hyperpolarize neuron (make more -)

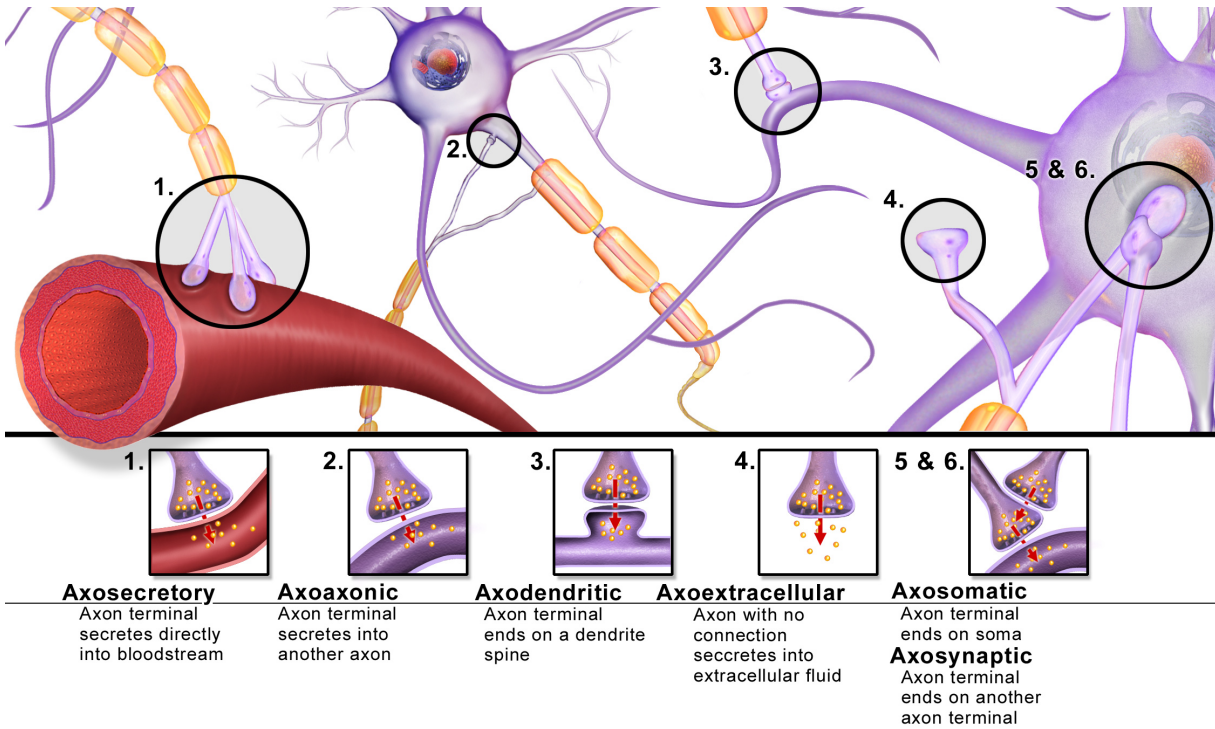
NTs inactivated

- Buffering
 - e.g., glutamate into astrocytes (Anderson & Swanson, 2000)
- *Reuptake via transporters* (https://en.wikipedia.org/wiki/Neurotransmitter_transporter)
 - e.g., serotonin via serotonin transporter (SERT)
- Enzymatic degradation
 - e.g., acetylcholine esterase (AChE) degrades acetylcholine (ACh)

Questions to ponder

- Why do NTs diffuse from pre- to post-synaptic membrane?
- Why must NTs be inactivated?
- What sort of PSP would *opening* a Na^+ channel produce?
- What sort of PSP would *opening* a Cl^- channel produce?
- What sort of PSP would *closing* a K^+ produce?

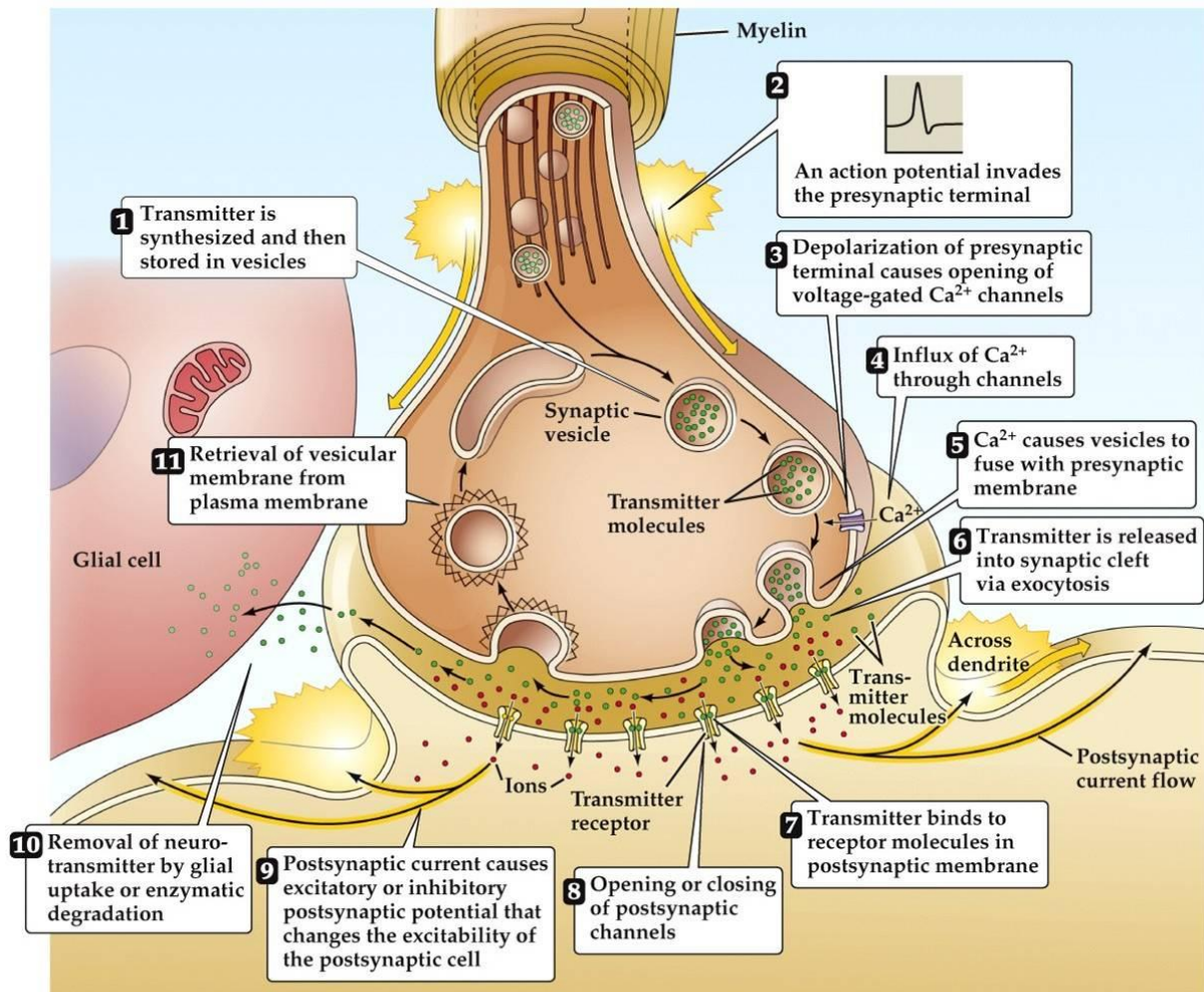
Synapse location and function



Source: Blausen.com staff

https://commons.wikimedia.org/wiki/File%3ABlausen_0843_SynapseTypes.png
 (https://commons.wikimedia.org/wiki/File%3ABlausen_0843_SynapseTypes.png)

- on dendrites
 - usually excitatory
- on cell bodies
 - usually inhibitory
- on axons
 - usually modulatory (change p(fire))



NEUROSCIENCE 5e, Figure 5.3

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Neurotransmitters

What are they?

- Chemicals produced by neurons
- Released by neurons
- Bound by neurons and other cells
- Send messages (have physiological effect on target cells)
- Inactivated after release

Things to know

- Neurotransmitter
- Where released from/to
- What receptor(s) bind it

Amino acids

Family	Neurotransmitter
Amino acids	Glutamate (https://en.wikipedia.org/wiki/Glutamate_(neurotransmitter)).
	γ aminobutyric acid (GABA) (https://en.wikipedia.org/wiki/Gamma-Aminobutyric_acid).
	Glycine
	Aspartate

Glutamate

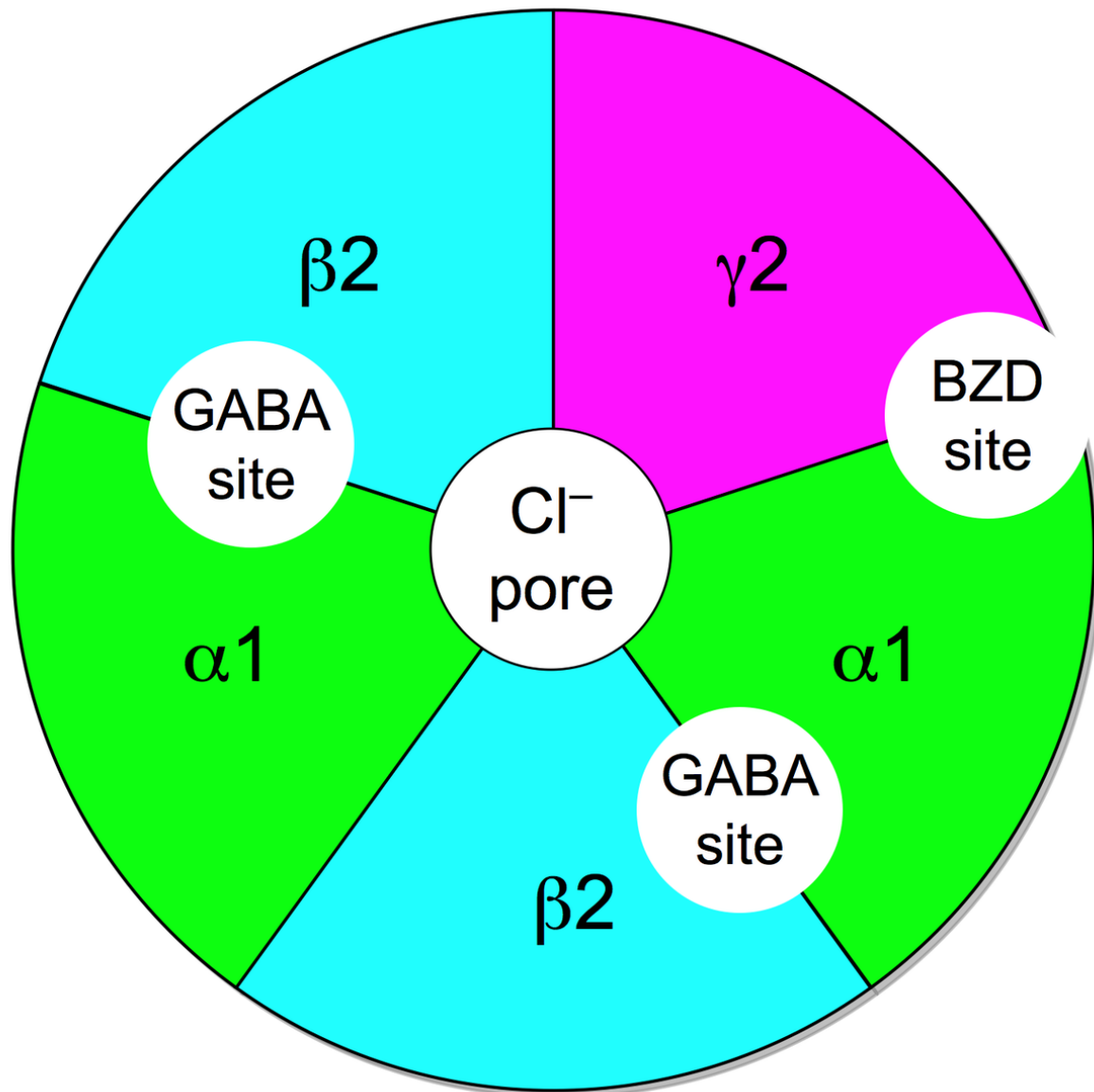
- Widespread in CNS (~ 1/2 all synapses)
- Primary excitatory NT in CNS
- Role in learning (via NMDA receptor)
- Receptors on neurons and glia (astrocytes and oligodendrocytes)
- Linked to umami (savory) taste sensation (think monosodium glutamate or MSG)
- Dysregulation in schizophrenia (McCutcheon, Krystal, & Howes, 2020) (<https://doi.org/10.1002/wps.20693>), mood disorders (Małgorzata, Paweł, Iwona, Brzostek, & Andrzej, 2020) (<http://dx.doi.org/10.1080/14728222.2020.1836160>)

Type	Receptor	Esp Permeable to
Ionotropic	AMPA	Na^+, K^+
	Kainate	
	NMDA	Ca^{++}
Metabotropic	mGlu	

γ aminobutyric acid (GABA)

- Primary inhibitory NT in CNS
- Excitatory in developing CNS, $[Cl^-]$ in \gg $[Cl^-]$ out
- Binding sites for benzodiazepines (BZD; e.g., Valium), barbiturates, ethanol, etc.
 - BZD affect subset of GABA-A receptors
 - Increase total Cl⁻ influx

Type	Receptor	Esp Permeable to
Ionotropic	GABA-A	Cl^-
Metabotropic	GABA-B	K^+



Source: <https://commons.wikimedia.org/wiki/File:GABAA-receptor-protein-example.png#/media/File:GABAA-receptor-protein-example.png>
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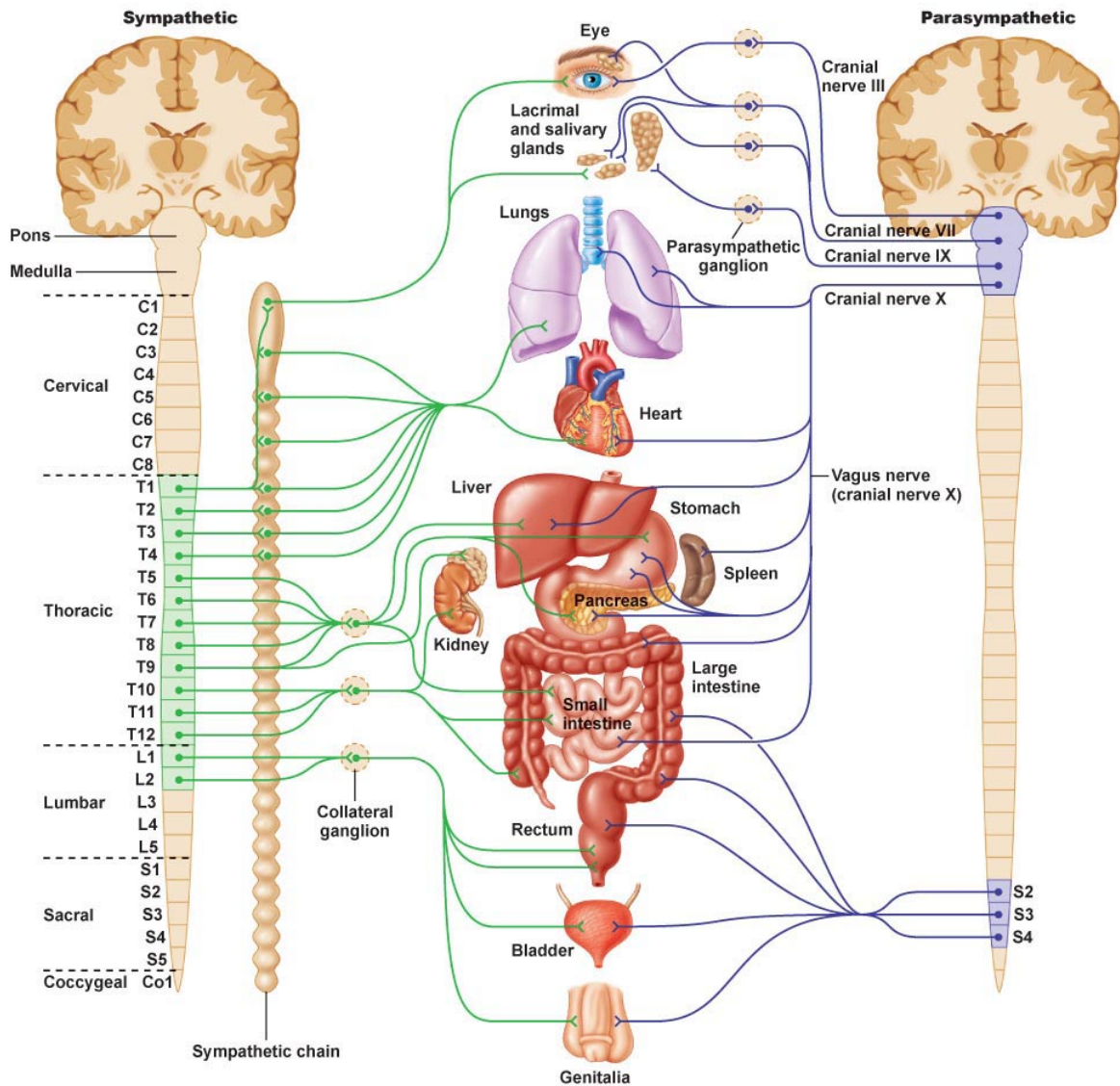
Other amino acid NTs

- *Aspartate*
 - Like Glu, stimulates NMDA receptor
- *Glycine*
 - Spinal cord interneurons

Acetylcholine (ACh)

- Primary excitatory NT of CNS output
- Somatic nervous system (motor neuron → neuromuscular junction)

- Autonomic nervous system (ANS)
 - Sympathetic branch: preganglionic neuron
 - Parasympathetic branch: pre/postganglionic



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Type	Receptor	Esp Permeable to	Blocked by
Ionotropic	Nicotinic (nAChR)	Na^+ , K^+	e.g., Curare
Metabotropic	Muscarinic (mAChR)	K^+	e.g., Atropine

Curare





Atropine

- aka, nightshade or belladonna
- inhibits (acts as an antagonist for) muscarinic ACh receptor



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Monoamine NTs

Family	Neurotransmitter	Comment
Monoamines	Dopamine (DA)	Catecholamine
	Norepinephrine (NE)/Noradrenaline (NAd)	Catecholamine
	Epinephrine (Epi)/Adrenaline (Ad)	Catecholamine

Serotonin (5-HT)

Indolamine

Melatonin

Indolamine

Histamine

- Synthesis pathway: DA -> NE/NAd -> Epi/Ad

Information processing

- Point-to-point
 - One sender, small number of recipients
 - Glu, GABA
- Broadcast
 - One sender, widespread recipients
 - DA, NE, 5-HT, melatonin, histamine
- Need to know
 - NT, where projecting, type of receptor to predict function

Dopamine

- Released by
 - Substantia nigra -> striatum, *meso-striatal projection*
 - Ventral tegmental area (VTA) -> nucleus accumbens, ventral striatum, hippocampus, amygdala, cortex; *meso-limbo-cortical projection*

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https://en.wikipedia.org/wiki/Dopaminergic_pathways
(https://en.wikipedia.org/wiki/Dopaminergic_pathways)

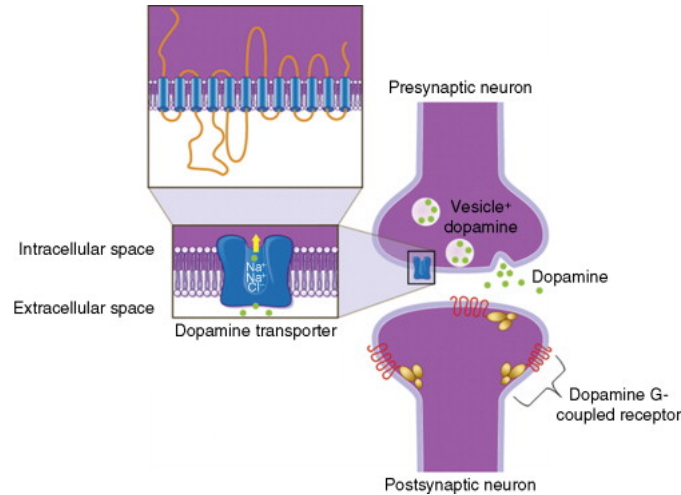
Clinical relevance

- Parkinson's Disease (mesostriatal)
 - DA agonists treat (agonists facilitate/increase transmission)
- ADHD (mesolimbocortical)
- Schizophrenia (mesolimbocortical)
 - DA antagonists treat
- Addiction (mesolimbocortical)

Inactivated via

- Chemical breakdown (e.g., via monoamine oxidase),
http://www.scholarpedia.org/article/Dopamine_anatomy#Dopamine_receptors
(http://www.scholarpedia.org/article/Dopamine_anatomy#Dopamine_receptors)
- Dopamine transporter (DAT)

- Psychostimulants (e.g., cocaine, methylphenidate) act upon. (“Dopamine transporter,” n.d.)
(<https://www.sciencedirect.com/topics/neuroscience/dopamine-transporter>)
- DAT also transports norepinephrine (NE)

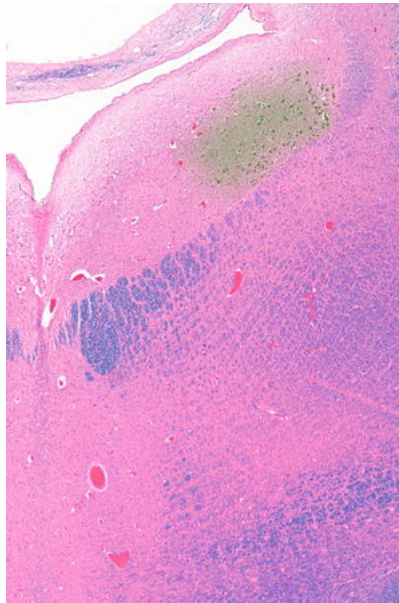


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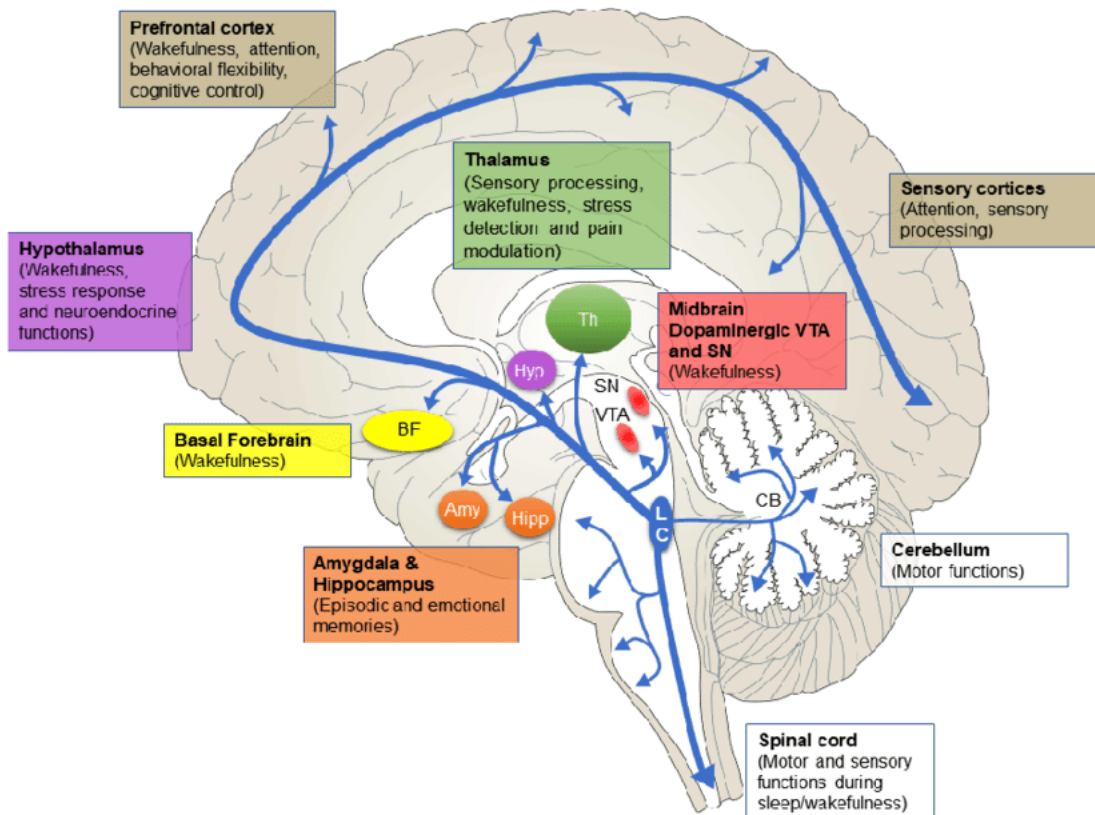
Type	Receptor	Comments
Metabotropic	D1-like (D1 and D5)	more prevalent
	D2-like (D2, D3, D4)	target of many antipsychotics

Norepinephrine

- Released by
 - locus coeruleus (http://www.scholarpedia.org/article/Locus_coeruleus) in pons/caudal tegmentum
 - postganglionic sympathetic neurons onto target tissues

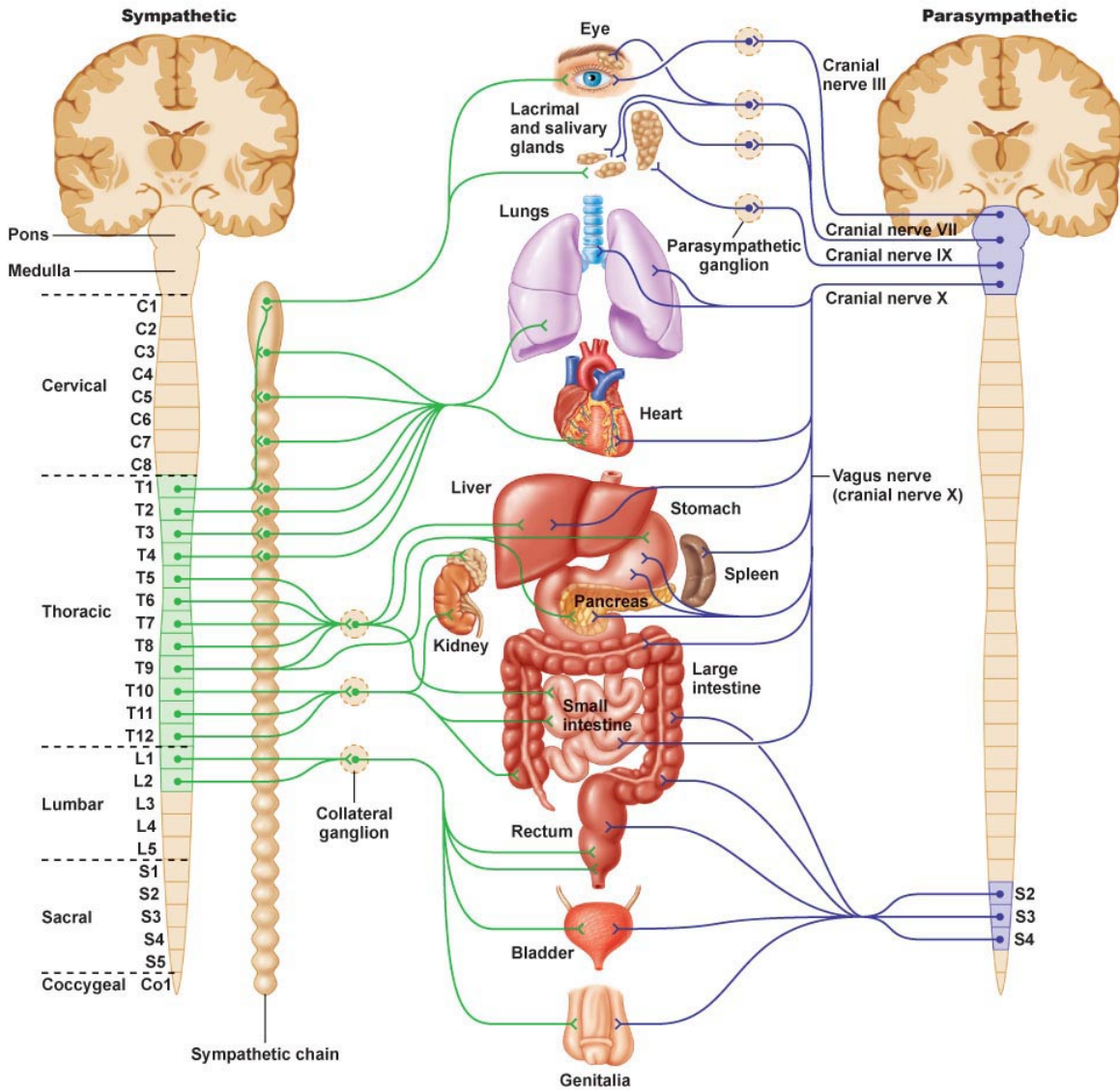


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- Role in arousal, mood, eating, sexual behavior

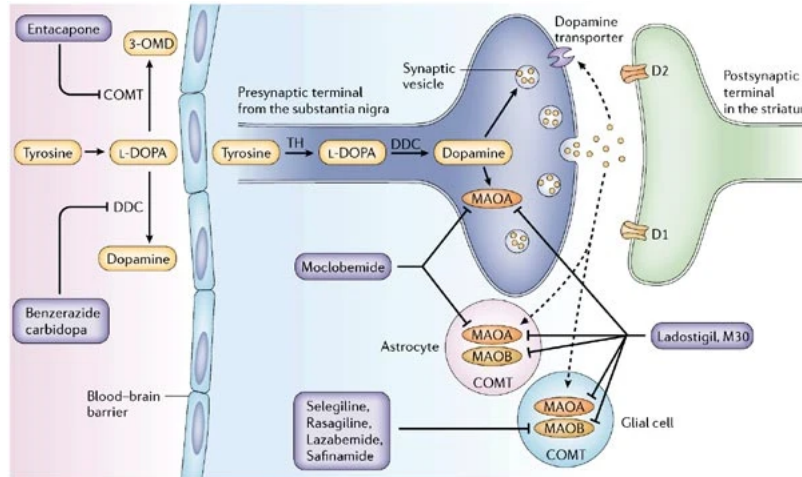
Clinical relevance

- ADHD, Alzheimer's Disease, Parkinson's Disease, depression

Inactivated by

- Norepinephrine transporter (NET), aka noradrenaline transporter (NAT)
 - Contributes to DA uptake, too.
- Also monoamine oxidase inhibitors (MAOIs)
 - inactivate monoamines in neurons, astrocytes

- MAOIs increase NE, DA
- Treatment for depression



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(Youdim, Edmondson, & Tipton, 2006) (<http://dx.doi.org/10.1038/nrn1883>)

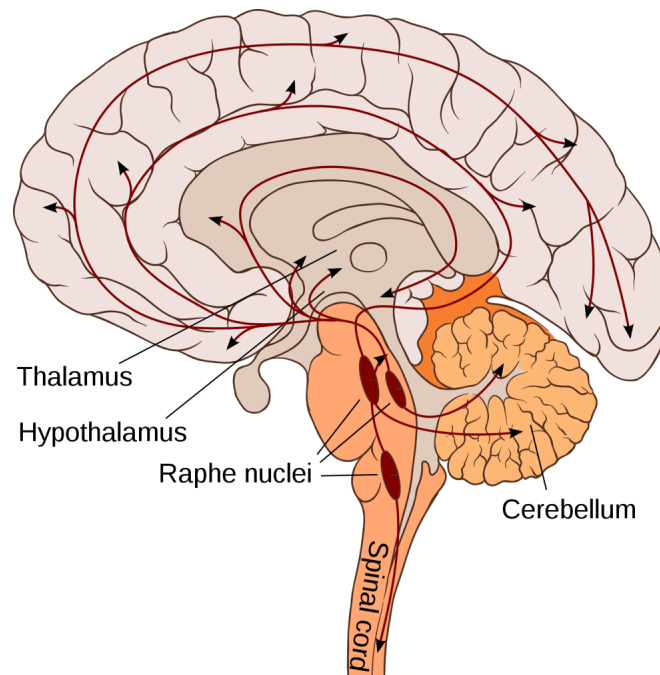
Type	Receptor	Comments
Metabotropic	α (1,2)	antagonists treat anxiety, panic
	β (1,2,3)	'beta blockers' in cardiac disease

Adrenaline/Epinephrine

- Synthesized from norepinephrine
- Both NT and hormone
 - As NT: Released in small amounts by medulla oblongata
 - As hormone: Released by adrenal medulla
- Binds to ($\alpha_{1,2}, \beta_{1,2,3}$ receptors in blood vessels, cardiac muscle, lungs, eye muscles controlling pupil dilation, liver, pancreas, etc.
- Release enhanced by cortisol from adrenal cortex
- Unusual in NOT being part of negative feedback system controlling its own release

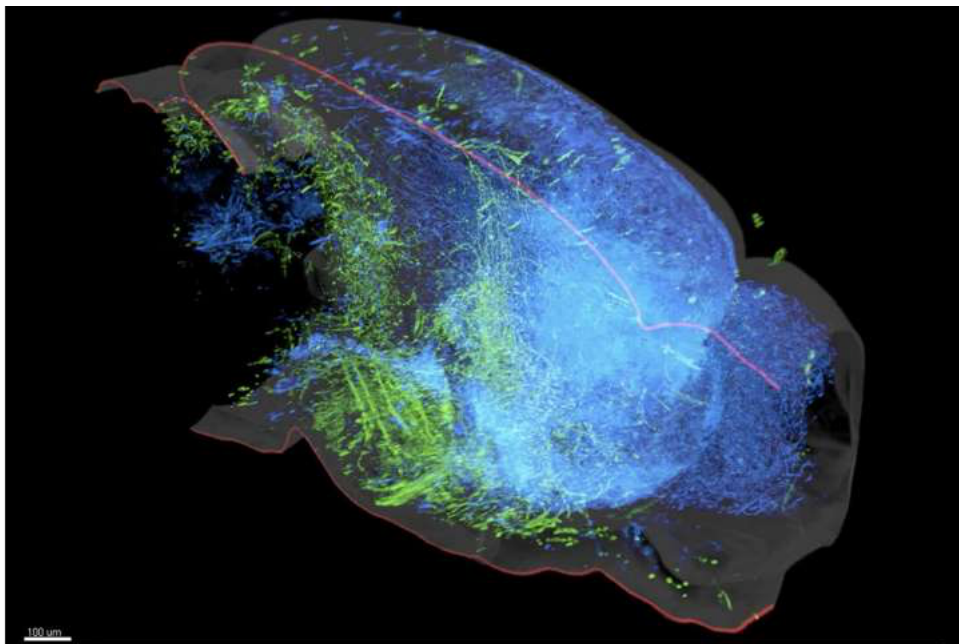
Serotonin (5-hydroxytryptamine or 5-HT)

- Released by *raphe nuclei* in brainstem



https://en.wikipedia.org/wiki/Serotonin_pathway
 (https://en.wikipedia.org/wiki/Serotonin_pathway)

- Role in mood, sleep, eating, pain, nausea, cognition, memory
- Modulates release of other NTs
- Most (90%; (De Ponti, 2004) (<http://dx.doi.org/10.1136/gut.2003.035568>)) of body's 5-HT regulates digestion
- Separate cortical, subcortical 5-HT projection pathways?



(Ren et al., 2018) (<http://dx.doi.org/10.1016/j.cell.2018.07.043>)

- Seven receptor families (5-HT 1-7) with 14 types

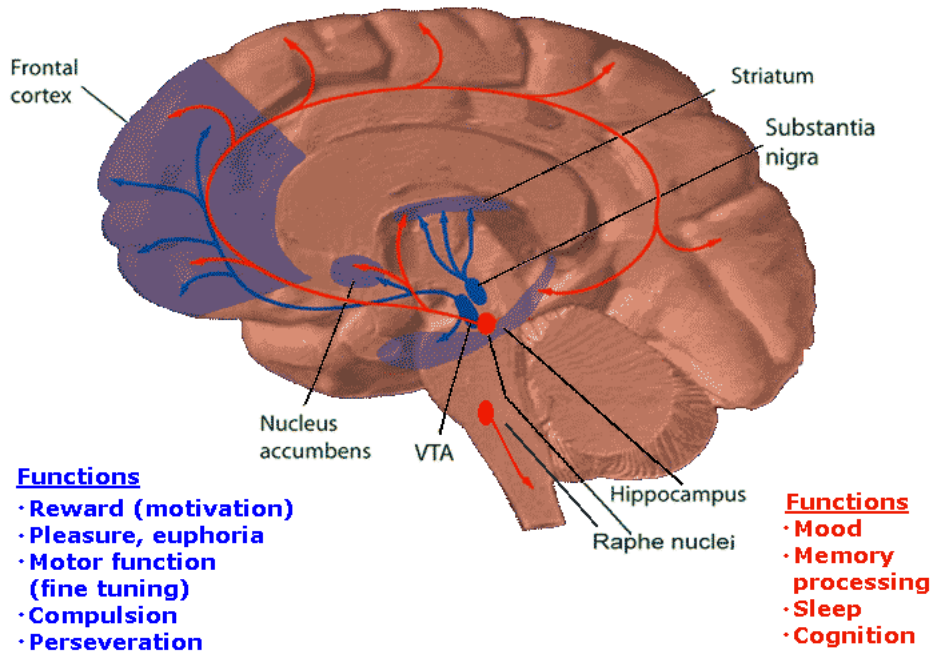
- All but one metabotropic

Clinical relevance

- Ecstasy (MDMA) disturbs serotonin
- So does LSD
- Fluoxetine (Prozac)
 - *Selective Serotonin Reuptake Inhibitor (SSRI)*
 - Treats depression, panic, eating disorders, others
- 5-HT₃ receptor antagonists are anti-mimetics used in treating nausea

Dopamine Pathways

Serotonin Pathways

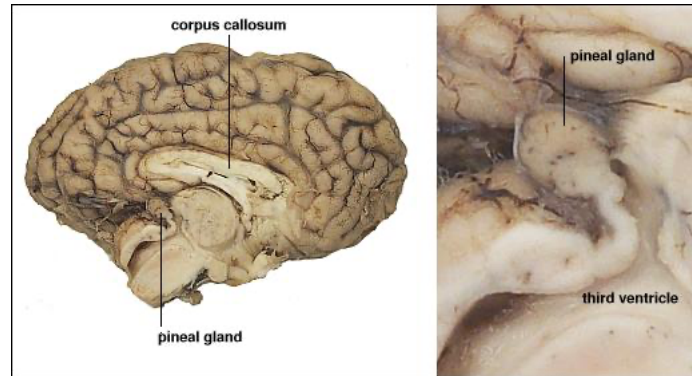


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Public Domain, Link (<https://commons.wikimedia.org/w/index.php?curid=45159949>)

- Different psychological roles (passive vs. active coping) associated with different 5-HT receptor subtypes? (Carhart-Harris & Nutt, 2017) (<http://dx.doi.org/10.1177/0269881117725915>)

Melatonin

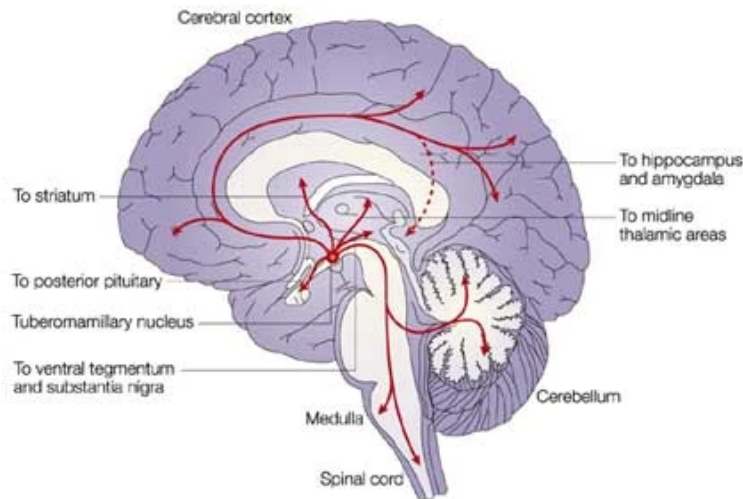
- Released by pineal gland (pine cone-like appearance)



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Histamine

- Released by hypothalamus, projects to whole brain



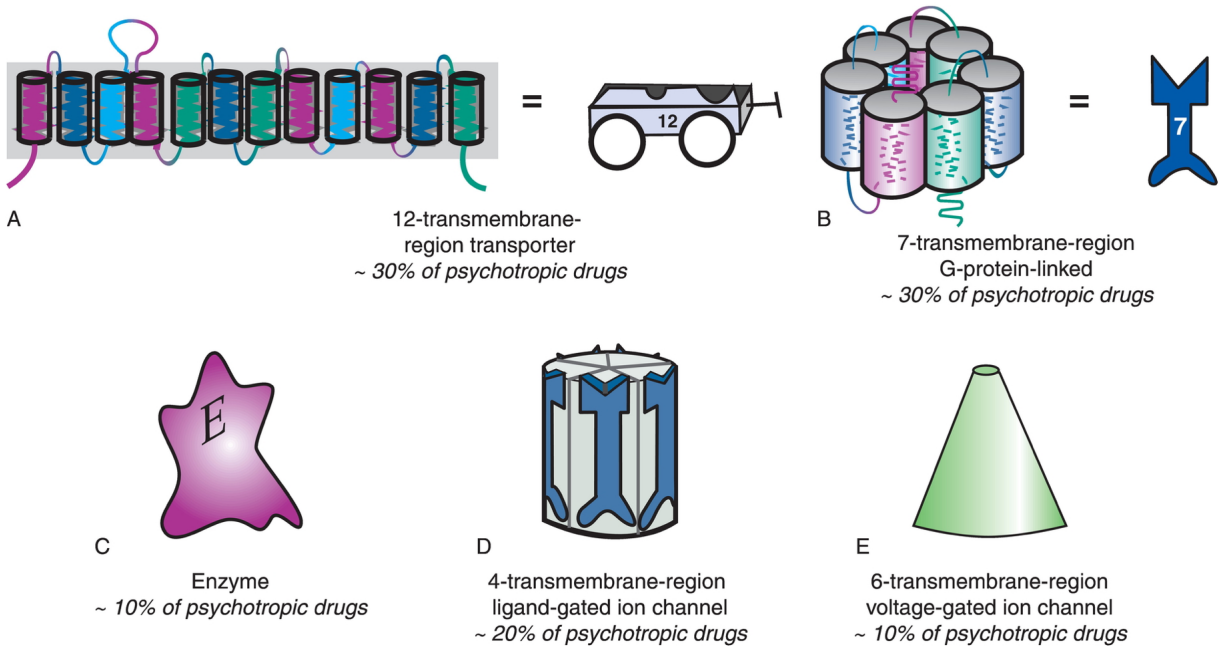
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<https://www.nature.com/articles/nrn1034> (<https://www.nature.com/articles/nrn1034>)

- H_1 - H_4 Metabotropic receptors, one ionotropic type in thalamus/hypothalamus
- Role in arousal/sleep regulation
- In body, part of immune/inflammatory response

Targets of psychotropic drugs

The Five Molecular Targets of Psychotropic Drugs



Source: https://stahlonline.cambridge.org/essential_4th_chapter.jsf?page=chapter2_summary.htm&name=Chapter%202&title=Summary
(https://stahlonline.cambridge.org/essential_4th_chapter.jsf?page=chapter2_summary.htm&name=Chapter%202&title=Summary)

Other NTs

- Gases
 - Nitric Oxide (NO), carbon monoxide (CO)
- Neuropeptides
 - Substance P and endorphins (endogenous morphine-like compounds) have role in pain
 - Orexin/hypocretin, project from lateral hypothalamus across brain, regulates appetite, arousal
 - Cholecystokinin (CCK) stimulates digestion
- Purines
 - Adenosine (inhibited by caffeine)
- Others
 - Anandamide (activates endogenous cannabinoid receptors)

Hormonal communication

- Chemicals secreted into blood
- Act on specific target tissues via receptors
- Produce specific effects

Examples of substances that are both hormones and NTs

- Melatonin
- Epinephrine/adrenaline
- Oxytocin
- Arginine Vasopressin (AVP) or Anti-Diuretic Hormone (ADH)

Behaviors under hormonal influence

Ingestive (eating/ drinking)

- Fluid levels
- Na, K, Ca levels
- Digestion
- Blood glucose levels

Reproduction-related

- Sexual Maturation
- Mating
- Birth
- Care giving

To threat/challenge

- Metabolism
- Heart rate, blood pressure
- Digestion
- Arousal

Common factors

- Biological imperatives
- Proscribed in space and time
- Foraging/hunting
 - Find targets distributed in space, evaluate, act upon
- Often involve others

Principles of hormonal action

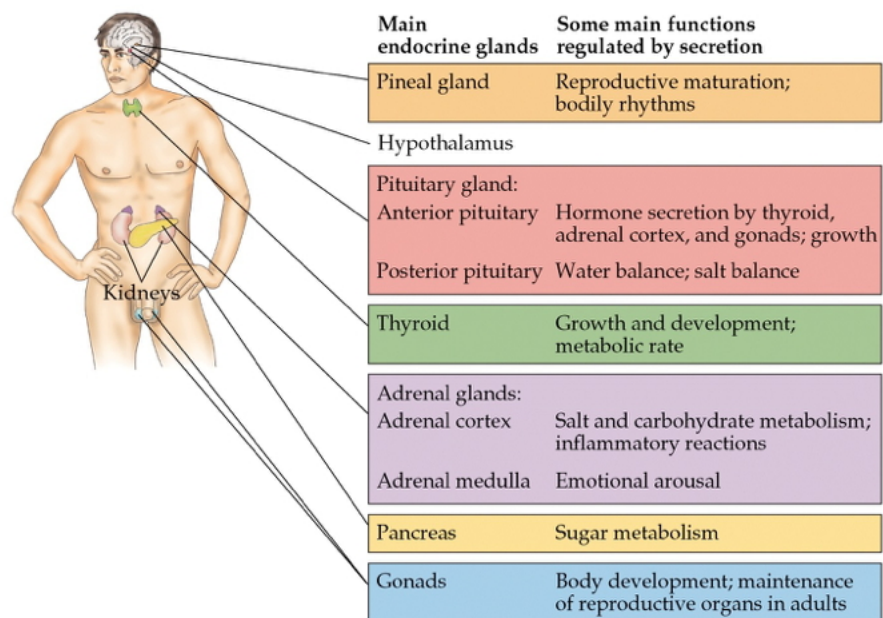
- Gradual action
- Change intensity or probability of behavior
- Behavior influences/influenced by hormones
 - +/- Feedback

- Multiple effects on different tissues
- Produced in small amounts; released in bursts
- Levels vary daily, seasonally
 - or are triggered by specific external/internal events
- Effect cellular metabolism
- Influence only cells with receptors
- Point to point vs. “broadcast”
 - Wider broadcast than neuromodulators

Similarities between neural and hormonal communication

- Chemical messengers stored for later release
- Release follows stimulation
- Action depends on specific receptors
- 2nd messenger systems common

Hormonal release sites



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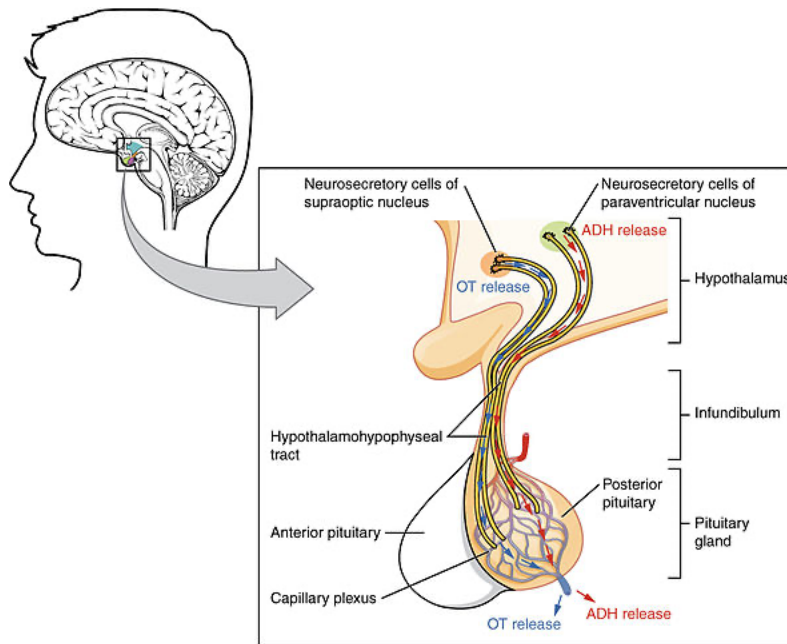
- CNS
 - Hypothalamus
 - Pituitary
 - Anterior
 - Posterior
 - Pineal gland
- Rest of body
 - Thyroid
 - Adrenal (*ad=adjacent, renal=kidney*) gland

- Adrenal cortex
- Adrenal medulla
- Gonads (testes/ovaries)

Two release systems from hypothalamus

Direct release

- Hypothalamus (paraventricular, supraoptic nucleus) to
- Posterior pituitary
 - Oxytocin
 - Arginine Vasopressin (AVP, vasopressin)



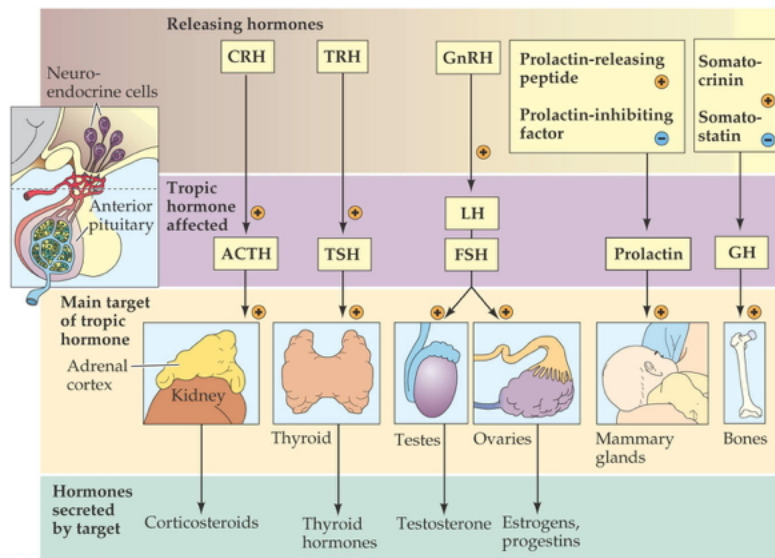
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Indirect release

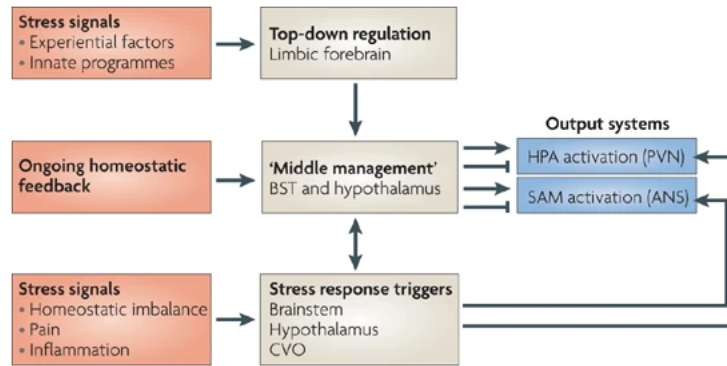
- Hypothalamus -> *releasing hormones*
- Anterior pituitary -> *tropic hormones*
- End organs



BIOLOGICAL PSYCHOLOGY, Fourth Edition, Figure 5.14 © 2004 Sinauer Associates, Inc.

Case studies

Responses to threat or challenge



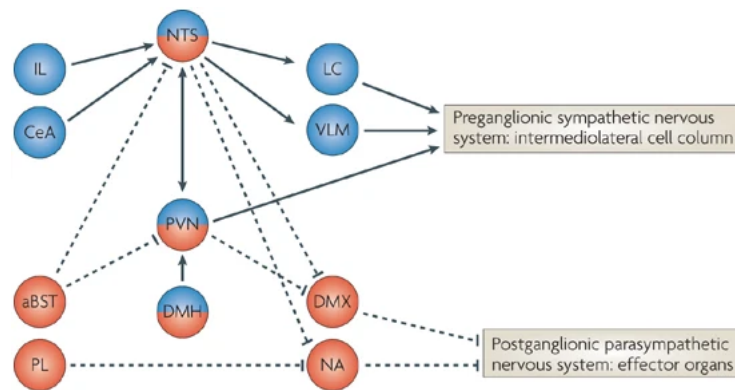
Nature Reviews | Neuroscience

(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

Figure 1: General scheme of brain acute-stress regulatory pathways. Stressors activate brainstem and/or forebrain limbic structures. The brainstem can generate rapid hypothalamic-pituitary-adrenal (HPA) axis and autonomic nervous system (ANS) responses through direct projections to hypophysiotrophic neurons in the paraventricular nucleus of the hypothalamus (PVN) or to preganglionic autonomic neurons (stress response triggers). By contrast, forebrain limbic regions have no direct connections with the HPA axis or the ANS and thus require intervening synapses before they can access autonomic or neuroendocrine neurons (top-down regulation). A high proportion of these intervening neurons are located in hypothalamic nuclei that are also responsive to homeostatic status, providing a mechanism by which the descending limbic information can be modulated according to the physiological status of the animal ('middle management'). BST, bed nucleus of the stria terminalis; CVO, circumventricular organ; SAM, sympathoadrenomedullary system.

(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

- Neural response
 - *Sympathetic Adrenal Medulla (SAM) response*
 - Sympathetic NS activation of adrenal medulla, other organs
 - Releases NE and Epi into bloodstream



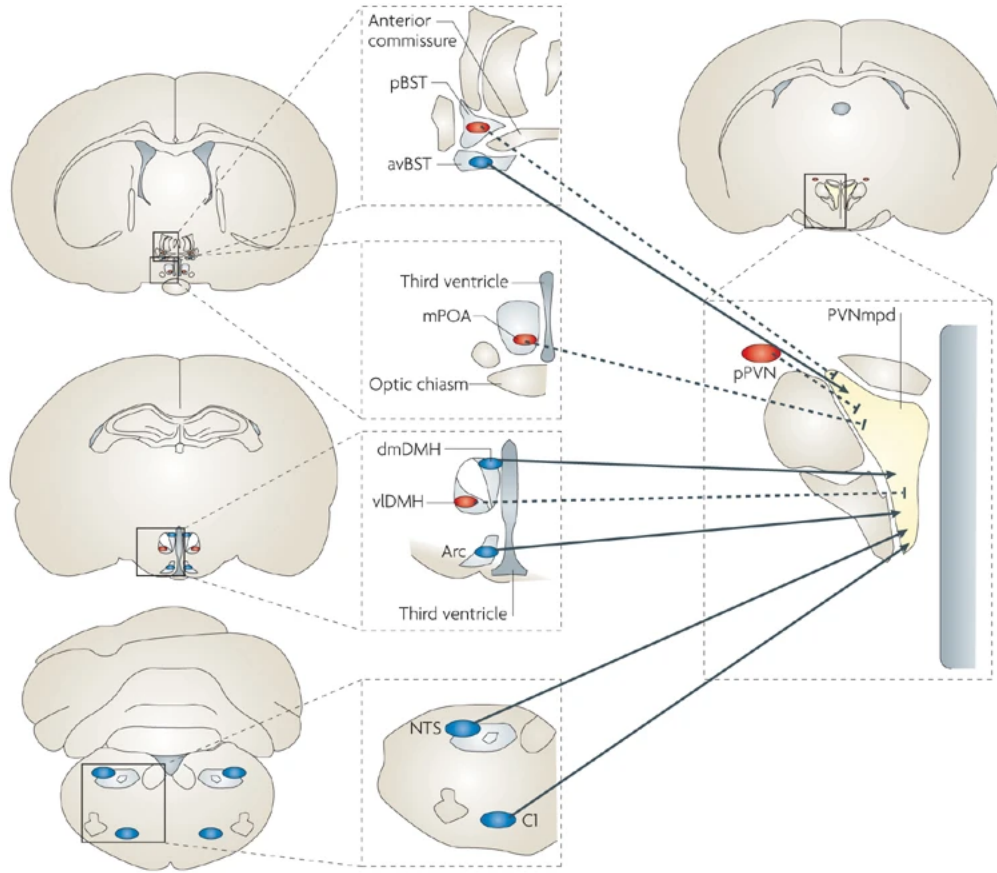
Nature Reviews | Neuroscience

(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

Figure 2: The brain circuitry that regulates autonomic stress responses. Stress-induced pre-autonomic outflow originates in multiple brain areas. The colours denote brain regions that are implicated in sympathetic activation (blue), parasympathetic activation (red) or both (bicoloured). The paraventricular nucleus of the hypothalamus (PVN) has substantial projections to both sympathetic and parasympathetic nuclei, including the nucleus of the solitary tract (NTS), the dorsal motor nucleus of the vagus nerve (DMX), the intermediolateral cell column (IML), the locus coeruleus (LC) and the ventrolateral medulla (VLM) (the latter two sets of projections are not shown for clarity). The rostral VLM, LC and PVN directly innervate the IML and are thought to initiate sympathetic responses. The NTS in turn receives direct input from neurons in the infralimbic cortex (IL), the central amygdala (CeA) and the PVN. Other hypothalamic regions, most notably the dorsomedial hypothalamus (DMH), modulate autonomic nervous system activation through connections with the PVN (and possibly other descending pathways) (see main text). Parasympathetic outflow is mediated largely by descending outputs from the DMX and the nucleus ambiguus (NA) and is under the direct influence of the prelimbic cortex (PL), the PVN and possibly other descending relays (see main text). Parasympathetic effects of the anterior bed nucleus of the stria terminalis (aBST) are probably mediated by relays in the PVN or the NTS. The anatomical complexity of autonomic nervous system integration is underscored by the mixing of sympathetic and parasympathetic projection neurons in individual nuclei.

(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

- Endocrine response
 - *Hypothalamic Pituitary Adrenal (HPA) axis*
 - Adrenal hormones released
- Hypothalamus
 - *Corticotropin Releasing Hormone (CRH)*
- Anterior pituitary
 - *Adrenocorticotrophic hormone (ACTH)*
- Adrenal cortex
 - *Glucocorticoids (e.g., cortisol)*
 - *Mineralocorticoids (e.g. aldosterone)*



Nature Reviews | Neuroscience

(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

Figure 3: The brain circuitry that regulates HPA axis stress responses. Stress-induced activation of the dorsal part of the medial parvocellular paraventricular nucleus of the hypothalamus (PVNmpd) originates in several brain regions (excitatory inputs are coloured blue with solid lines and inhibitory (GABA (γ -aminobutyric acid)-ergic) inputs are coloured red with dashed lines). The paraventricular nucleus of the hypothalamus (PVN) receives direct noradrenergic, adrenergic and peptidergic innervation from the nucleus of the solitary tract (NTS). The dorsomedial components of the dorsomedial hypothalamus (dmDMH) and the arcuate nucleus (Arc) provide intrahypothalamic stress excitation. The anterior part of the bed nucleus of the stria terminalis (BST), particularly the anteroventral nucleus of the BST (avBST), activates hypothalamic-pituitary-adrenocortical (HPA) axis stress responses. The PVN also receives a stress-excitatory drive from the dorsal raphe, the tuberomammillary nucleus, the supramammillary nucleus and the spinal cord, among others (omitted in the interest of space). Activation of the PVNmpd is inhibited by numerous hypothalamic circuits, including the medial preoptic area (mPOA), the ventrolateral component of the dorsomedial hypothalamus (vlDMH) and local neurons in the peri-PVN region (pPVN), encompassing the PVN surround and the subparaventricular zone. The posterior subregions of the bed nucleus of the stria terminalis (pBST) provide a prominent forebrain inhibition of HPA axis responses; most of these inputs are GABAergic. Brain sections are modified, with permission, from Ref. 154 © (1998) Academic Press.

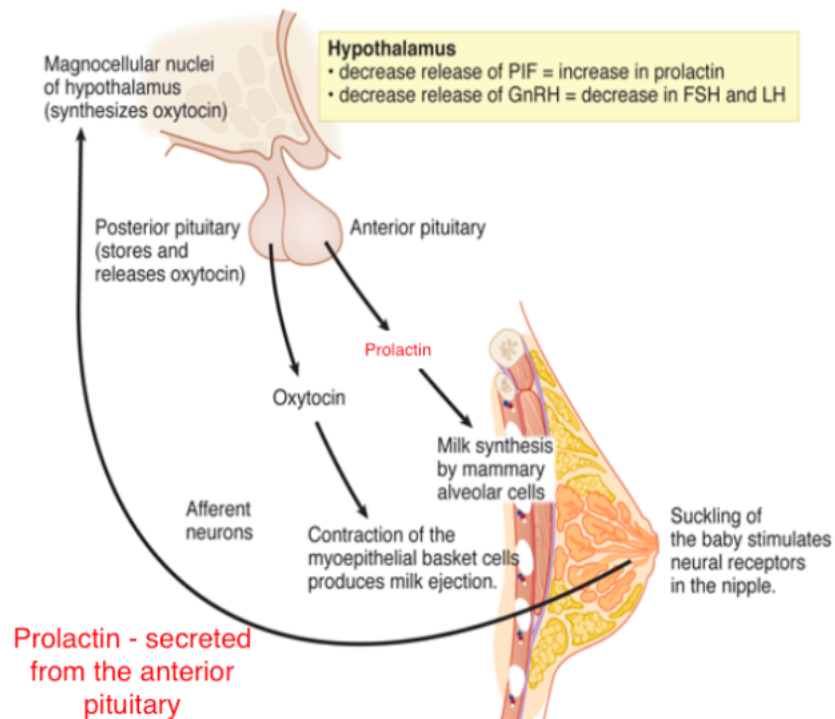
(Ulrich-Lai & Herman, 2009) (<http://doi.org/10.1038/nrn2647>)

Adrenal hormones

- *Steroids*
 - Derived from cholesterol
- *Cortisol*
 - increases blood glucose, anti-inflammatory effects
 - negative consequences of prolonged exposure
- *Aldosterone*
 - Regulates Na (and water)

Reproductive behavior – the milk letdown reflex

- Supraoptic & Paraventricular nucleus (PVN) of hypothalamus releases oxytocin
 - Into bloodstream via posterior pituitary (endocrine)
 - Onto neurons in nucleus accumbens (neurocrine), amygdala, brainstem
- Oxytocin stimulates milk ducts to secrete



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Oxytocin's role...

- Sexual arousal
- Released in bursts during orgasm
- Stimulates uterine, vaginal contraction during labor
 - But mouse OXY knock-out model still engages in reproductive behavior and gives birth without incident.
- Oxytocin cells in ovarian corpus luteum, testicles, retina, adrenal medulla, pancreas
- Links to social interaction, bonding (Weisman & Feldman, 2013)
 (http://dx.doi.org/10.1016/j.biopsych.2013.05.026)
- Alters face processing in autism (Domes et al., 2013)
 (http://dx.doi.org/10.1016/j.biopsych.2013.02.007)
- May inhibit fear/anxiety-related behaviors by gating amygdala (Viviani et al., 2011)
 (http://doi.org/10.1126/science.1201043)

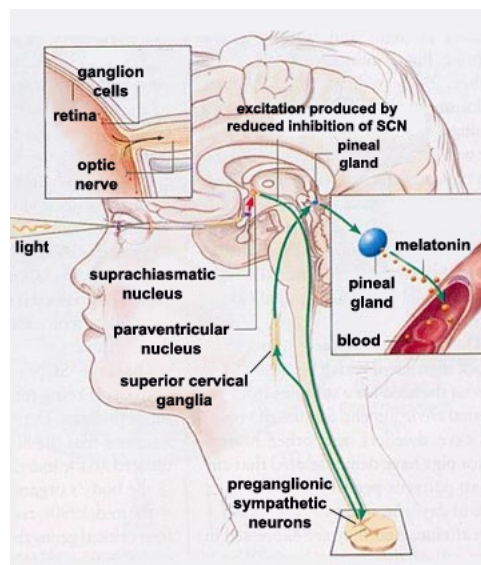


([http://columbusfreepress.com/sites/default/files/keep-calm-and-release-oxytocin-2\(1\).png](http://columbusfreepress.com/sites/default/files/keep-calm-and-release-oxytocin-2(1).png))

Circadian rhythms

Melatonin

- Diurnal rhythm
- Night time peak, early morning low
- Secretion suppressed by short wavelength or “blue” light (< 460-480 nm)
- Rhythm irregular until ~3 mos post-natal (Ardura, Gutierrez, Andres, & Agapito, 2003) (<http://dx.doi.org/68571>)
- Peak weakens, broadens with age



- Pathway
 - Suprachiasmatic nucleus (SCN) of the hypothalamus
 - Paraventricular nucleus of the hypothalamus
 - Spinal cord
 - Superior cervical ganglion
 - Pineal gland



Thinking about neurochemical influences

- Measure hormones in blood, saliva; can't effectively measure NTs
- Multivariate, nonlinear, mutually interacting
- Varied time scales
 - Phasic (e.g., cortisol in response to challenge)
 - Periodic (e.g., melatonin; diurnal cortisol)
- Peripheral effects + neural feedback
- State variables and behavior
 - Are your participants sleepy, hungry, horny, distressed...
 - Endogenous & exogenous influences
 - Systems interact; need better, broader, and denser measurement

Enteric nervous system

Anatomy

- A component of the Autonomic Nervous System (ANS), along with the Sympathetic (SNS) and Parasympathetic Nervous Systems (PNS).
- number of neurons comparable to entire spinal cord (Alloway & Pritchard)

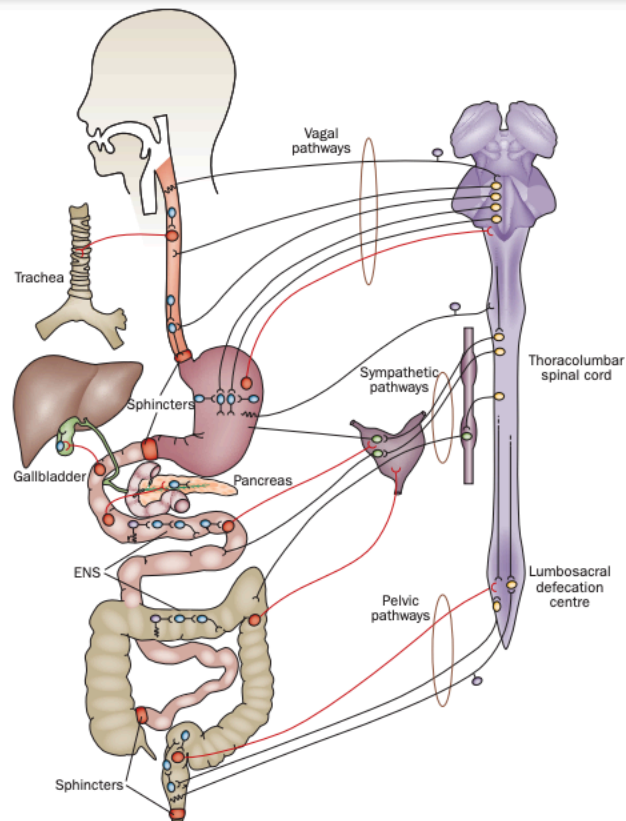
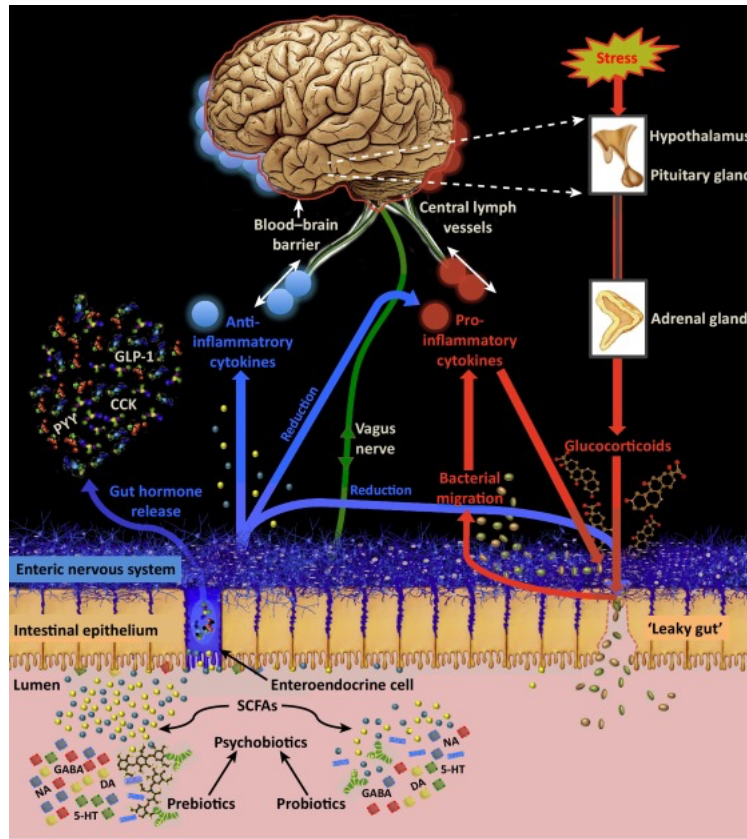


Figure 1 | The innervation of the gastrointestinal tract. The neural connections between the ENS and CNS, and neural connections between gastrointestinal organs, are quite different from those depicted in textbooks. The digestive system contains full reflex circuits of the ENS (motor neurons and interneurons in blue, sensory neurons in purple). Pathways from the gastrointestinal tract project outwards, via intestinofugal neurons (red), to the CNS (neurons in yellow), sympathetic ganglia, gallbladder and pancreas. Neurons in sympathetic prevertebral ganglia (green) receive both CNS and ENS inputs. Sensory information goes both to the ENS, via intrinsic primary afferent (sensory) neurons (purple) and to the CNS via extrinsic primary afferent neurons (also purple) that follow spinal and vagal afferent routes. Pathways from the CNS reach the ENS and gastrointestinal effector tissues through vagal, sympathetic and pelvic pathways. Abbreviations: CNS, central nervous system; ENS, enteric nervous system.

(Furness, 2012) (<http://dx.doi.org/10.1038/nrgastro.2012.32>)

Gut/brain connection



Trends in Neurosciences

(Sarkar et al., 2016) (<https://doi.org/10.1016/j.tins.2016.09.002>)

Figure 1. Systems-Level Overview of Psychobiotic Action. Blue arrows indicate psychobiotic processes and effects, while red arrows indicate processes associated with leaky gut and inflammation. Probiotics directly introduce beneficial bacteria such as Lactobacilli and Bifidobacteria into the gut. Prebiotics (e.g., galacto-oligosaccharides) support the growth of such bacteria. SCFAs and gut hormones: Both probiotics and prebiotics increase production of short-chain fatty acids (SCFAs), which interact with gut mucosal enteroendocrine cells and catalyse the release of gut hormones such as cholecystinin (CCK), peptide tyrosine tyrosine (PYY) and glucagon-like peptide- 1 (GLP-1). Prebiotics may have stronger effects in this regard in comparison to probiotics. SCFAs and gut hormones enter circulation and can migrate into the central nervous system. Gut hormones are also secreted by tissues other than enteroendocrine cells. Neurotransmitters: psychobiotics enhance neurotransmitter production in the gut, including dopamine (DA), serotonin (5-HT), noradrenaline (NA), and γ -aminobutyric acid (GABA), which likely modulate neurotransmission in the proximal synapses of the enteric nervous system. Vagal connections: the vagus nerve synapses on enteric neurons and enables gut–brain communication. Stress, barrier function, and cytokines: barrier dysfunction is exacerbated through stress-induced glucocorticoid exposure. This enables migration of bacteria with pro-inflammatory components, increasing inflammation directly and also triggering a rise in pro-inflammatory cytokines via the immunogenic response. These cytokines impair the integrity of the blood–brain barrier and permit access to potentially pathogenic or inflammatory elements. Pro-inflammatory cytokines (red circles) also reduce the integrity of the gut barrier. Psychobiotic action restores gut barrier function and decreases circulating concentrations of glucocorticoids and pro-inflammatory cytokines. They also increase concentrations of anti-inflammatory cytokines (blue circles), which enhance integrity of the blood–brain barrier, the gut barrier, and reduce overall inflammation. Cytokines clustering at the brain represent cytokine interaction with the blood–brain barrier. Central lymphatic vessels: cytokines may interact more directly with the brain than previously appreciated through the recently discovered central lymphatic vessels.

(Sarkar et al., 2016) (<https://doi.org/10.1016/j.tins.2016.09.002>)

- “Moody microbes or fecal phrenology?” (Forsythe, Kunze, & Bienenstock, 2016) (<http://dx.doi.org/10.1186/s12916-016-0604-8>).
- 2015 systematic review (with only $n = 10$ trials) shows “very limited evidence for the efficacy of probiotic interventions in psychological outcomes.” (Romijn & Rucklidge, 2015) (<http://dx.doi.org/10.1093/nutrit/nuv025>).
- ROG: But do we have good data about what people actually consume?

Physiology

- electrogastrogram (EGG) from ENS (Al Tae & Al-Jumaily, 2020) (http://dx.doi.org/10.1007/978-3-030-14347-3_50) measures gastric motility

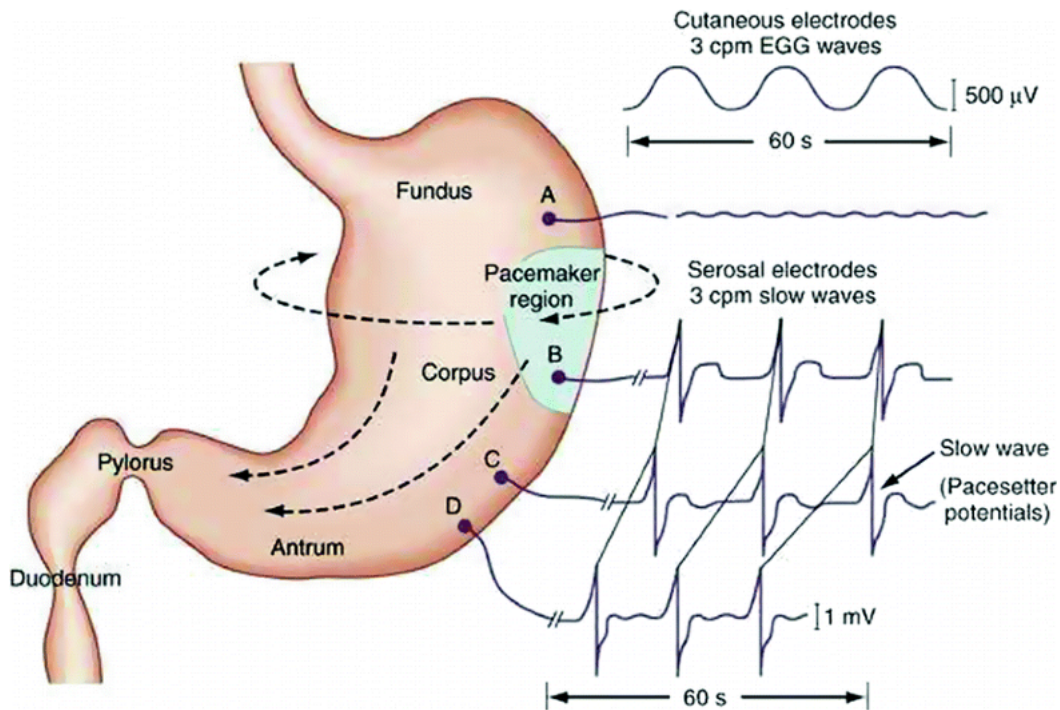
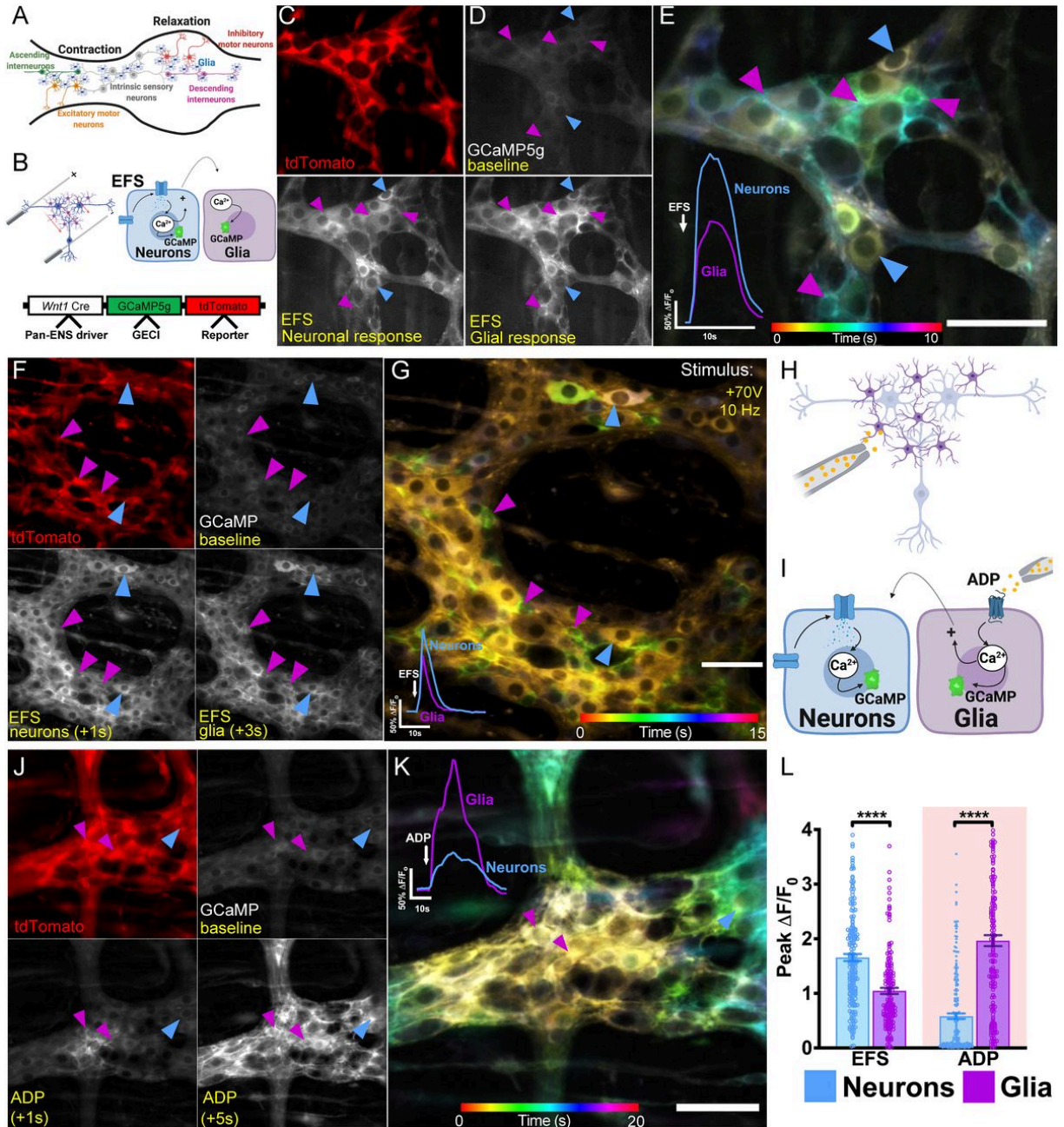


Fig. 1. Gastric pacesetter potentials or slow waves originate from the pacemaker area on the greater curve. Pacesetter potentials travel in a circumferential and aboral direction at a rate of approximately 3 cycles per minute (cpm). The cutaneously recorded electrogastrogram shows 3-cpm wave pattern. The fundus has no rhythmic electrical activity.

(Al Tae & Al-Jumaily, 2020) (http://dx.doi.org/10.1007/978-3-030-14347-3_50)

- glial cells and neurons work together (Ahmadzai, Seguella, & Gulbransen, 2021) (<http://dx.doi.org/10.1073/pnas.2025938118>)



(Ahmadzai, Seguela, & Gulbransen, 2021) (<http://dx.doi.org/10.1073/pnas.2025938118>)

References

- Ahmadzai, M. M., Seguela, L., & Gulbransen, B. D. (2021). Circuit-specific enteric glia regulate intestinal motor neurocircuits. *Proceedings of the National Academy of Sciences of the United States of America*, 118(40). <https://doi.org/10.1073/pnas.2025938118> (<https://doi.org/10.1073/pnas.2025938118>)
- Al Tae, A., & Al-Jumaily, A. (2020). Electrogastrogram based medical applications an overview and processing frame work. In *Hybrid intelligent systems* (pp. 511–520). Springer International Publishing. https://doi.org/10.1007/978-3-030-14347-3_50

- (https://doi.org/10.1007/978-3-030-14347-3_50)
- Anderson, C. M., & Swanson, R. A. (2000). Astrocyte glutamate transport: Review of properties, regulation, and physiological functions. *Glia*, 32(1), 1–14.
[https://doi.org/10.1002/1098-1136\(200010\)32:1<1::AID-GLIA10>3.0.CO;2-W](https://doi.org/10.1002/1098-1136(200010)32:1<1::AID-GLIA10>3.0.CO;2-W)
 ([https://doi.org/10.1002/1098-1136\(200010\)32:1<1::AID-GLIA10>3.0.CO;2-W](https://doi.org/10.1002/1098-1136(200010)32:1<1::AID-GLIA10>3.0.CO;2-W))
- Ardura, J., Gutierrez, R., Andres, J., & Agapito, T. (2003). Emergence and evolution of the circadian rhythm of melatonin in children. *Horm. Res.*, 59(2), 66–72.
<https://doi.org/68571> (<https://doi.org/68571>)
- Carhart-Harris, R. L., & Nutt, D. J. (2017). Serotonin and brain function: A tale of two receptors. *Journal of Psychopharmacology*, 31(9), 1091–1120.
<https://doi.org/10.1177/0269881117725915>
 (<https://doi.org/10.1177/0269881117725915>)
- De Ponti, F. (2004). Pharmacology of serotonin: What a clinician should know. *Gut*, 53(10), 1520–1535. <https://doi.org/10.1136/gut.2003.035568>
 (<https://doi.org/10.1136/gut.2003.035568>)
- Domes, G., Heinrichs, M., Kumbier, E., Grossmann, A., Hauenstein, K., & Herpertz, S. C. (2013). Effects of intranasal oxytocin on the neural basis of face processing in autism spectrum disorder. *Biological Psychiatry*, 74(3), 164–171.
<https://doi.org/http://dx.doi.org/10.1016/j.biopsych.2013.02.007>
 (<http://dx.doi.org/10.1016/j.biopsych.2013.02.007>)
- Dopamine transporter. (n.d.).
<https://www.sciencedirect.com/topics/neuroscience/dopamine-transporter>
 (<https://www.sciencedirect.com/topics/neuroscience/dopamine-transporter>).
 Retrieved from <https://www.sciencedirect.com/topics/neuroscience/dopamine-transporter> (<https://www.sciencedirect.com/topics/neuroscience/dopamine-transporter>)
- Forsythe, P., Kunze, W., & Bienenstock, J. (2016). Moody microbes or fecal phrenology: What do we know about the microbiota-gut-brain axis? *BMC Medicine*, 14, 58.
<https://doi.org/10.1186/s12916-016-0604-8> (<https://doi.org/10.1186/s12916-016-0604-8>)
- Furness, J. B. (2012). The enteric nervous system and neurogastroenterology. *Nature Reviews. Gastroenterology & Hepatology*, 9(5), 286–294.
<https://doi.org/10.1038/nrgastro.2012.32> (<https://doi.org/10.1038/nrgastro.2012.32>)
- Hastoy, B., Clark, A., Rorsman, P., & Lang, J. (2017). Fusion pore in exocytosis: More than an exit gate? A β -cell perspective. *Cell Calcium*, 68, 45–61.
<https://doi.org/10.1016/j.ceca.2017.10.005> (<https://doi.org/10.1016/j.ceca.2017.10.005>)
- Haucke, V., Neher, E., & Sigrist, S. J. (2011). Protein scaffolds in the coupling of synaptic exocytosis and endocytosis. *Nature Reviews. Neuroscience*, 12(3), 127–138.
<https://doi.org/10.1038/nrn2948> (<https://doi.org/10.1038/nrn2948>)
- Małgorzata, P., Paweł, K., Iwona, M. L., Brzostek, T., & Andrzej, P. (2020). Glutamatergic

- dysregulation in mood disorders: Opportunities for the discovery of novel drug targets. *Expert Opinion on Therapeutic Targets*, 24(12), 1187–1209.
<https://doi.org/10.1080/14728222.2020.1836160>
(<https://doi.org/10.1080/14728222.2020.1836160>)
- McCutcheon, R. A., Krystal, J. H., & Howes, O. D. (2020). Dopamine and glutamate in schizophrenia: Biology, symptoms and treatment. *World Psychiatry: Official Journal of the World Psychiatric Association*, 19(1), 15–33. <https://doi.org/10.1002/wps.20693>
(<https://doi.org/10.1002/wps.20693>)
- Ren, J., Friedmann, D., Xiong, J., Liu, C. D., Ferguson, B. R., Weerakkody, T., ... Luo, L. (2018). Anatomically defined and functionally distinct dorsal raphe serotonin sub-systems. *Cell*. <https://doi.org/10.1016/j.cell.2018.07.043>
(<https://doi.org/10.1016/j.cell.2018.07.043>)
- Romijn, A. R., & Rucklidge, J. J. (2015). Systematic review of evidence to support the theory of psychobiotics. *Nutrition Reviews*, 73(10), 675–693.
<https://doi.org/10.1093/nutrit/nuv025> (<https://doi.org/10.1093/nutrit/nuv025>)
- Sarkar, A., Lehto, S. M., Harty, S., Dinan, T. G., Cryan, J. F., & Burnet, P. W. J. (2016). Psychobiotics and the manipulation of Bacteria–Gut–Brain signals. *Trends in Neurosciences*, 39(11), 763–781. <https://doi.org/10.1016/j.tins.2016.09.002>
(<https://doi.org/10.1016/j.tins.2016.09.002>)
- Ulrich-Lai, Y. M., & Herman, J. P. (2009). Neural regulation of endocrine and autonomic stress responses. *Nature Reviews Neuroscience*, 10(6), 397–409.
<https://doi.org/10.1038/nrn2647> (<https://doi.org/10.1038/nrn2647>)
- Viviani, D., Charlet, A., Burg, E. van den, Robinet, C., Hurni, N., Abatis, M., ... Stoop, R. (2011). Oxytocin selectively gates fear responses through distinct outputs from the central amygdala. *Science*, 333(6038), 104–107.
<https://doi.org/10.1126/science.1201043> (<https://doi.org/10.1126/science.1201043>)
- Weisman, O., & Feldman, R. (2013). Oxytocin effects on the human brain: Findings, questions, and future directions. *Biological Psychiatry*, 74(3), 158–159.
<https://doi.org/http://dx.doi.org/10.1016/j.biopsych.2013.05.026>
(<http://dx.doi.org/10.1016/j.biopsych.2013.05.026>)
- Youdim, M. B. H., Edmondson, D., & Tipton, K. F. (2006). The therapeutic potential of monoamine oxidase inhibitors. *Nature Reviews. Neuroscience*, 7(4), 295–309.
<https://doi.org/10.1038/nrn1883> (<https://doi.org/10.1038/nrn1883>)